ELSEVIER

Contents lists available at ScienceDirect

Bioresource Technology

journal homepage: www.elsevier.com/locate/biortech



Bioaerosols associated with animal production operations

Patricia D. Millner*

United States Department of Agriculture, Agricultural Research Service, EMFSL, Beltsville, MD 20705, USA

ARTICLE INFO

Article history: Received 1 July 2008 Received in revised form 5 March 2009 Accepted 10 March 2009 Available online 22 April 2009

Keywords: Manure Air dispersion Pathogens Organic dust Odor

ABSTRACT

Air emissions from animal housing and manure management operations include a complex mixture of biological, microbial, and inorganic particulates along with odorous volatile compounds. This report highlights the state of current issues, technical knowledge, and remaining challenges to be addressed in evaluating the impacts of airborne microorganisms, dusts, and odorants on animals and workers at animal production facilities and nearby communities. Reports documenting bioaerosol measurements illustrate some of the technical issues related to sample collection, analysis, as well as dispersion and transport to off-farm locations. Approaches to analysis, mitigation and modeling transport are discussed in the context of the risk reduction and management of airborne spread of bioaerosols from animal operations. The need for standardization and validation of bioaerosol collection and analytical techniques for indoor as well as outdoor animal agriculture settings is critical to evaluation of health effects from modern animal production systems that are increasingly situated near communities.

Published by Elsevier Ltd.

1. Introduction

The size and geospatial distribution of livestock and poultry operations continues to respond to demographics, land pressure, and economic development. Existing trends toward intensification and industrialization of meat animal production are projected to increase globally. Microbes of concern in animal production have been studied by veterinary, public health, sanitation and agricultural scientists (Smith et al., 2005; Strauch and Ballarini, 1994). Animal confinement tends to increase the overall microbial load in the immediate production environment by virtue of the increased volumes of feed, animals, and organic residuals (manure and wastewater) present, and the increased handling and management required. With more animals, materials, equipment, and workers in the production facility, there is a concurrent increase in three distinct yet interrelated types of airborne materials: (1) bioaerosols, (2) dust (mineral particulates serving as carriers), and (3) odorous volatile compounds.

Animal housing typically exposes animals and workers to substantial concentrations of volatile compounds (NH₃, CH₄, numerous organics, and H₂S), dust (fine particulates, endotoxin, animal dander, animal feed and excreta), and a variety of bioaerosols (bacteria, endotoxin, viruses, parasites, fungi, mycotoxin, insect parts, pollen, and grain particles) that can have adverse health effects (Clark et al., 1983; Cole et al., 2000; Douwes et al., 2003; Nowak, 1998; Zejda et al., 1994). Bioaerosols initially generated indoors may disperse outdoors. Manure application to fields may generate

bioaerosols, dust, and odors that transit to and beyond a property boundary. The impacts that new technologies to mitigate biological, nutrient, or odorant concentrations in animal production facilities, have concurrently on bioaerosols, dust, and odorant compounds will continue to require evaluation.

This report highlights the state of current issues, technical knowledge, and remaining challenges to be addressed in evaluating the relationships among airborne microorganisms, dusts, and odorants on animals and workers at animal production facilities and nearby communities. Reports documenting bioaerosol measurements illustrate some of the technical issues related to sample collection, analysis, as well as dispersion and transport to off-farm locations. Approaches to analysis, mitigation and modeling transport are discussed in the context of the risk reduction and management of airborne spread of bioaerosols from animal operations.

2. Bioaerosols, dust, and odor relationships

2.1. Bioaerosols

Bioaerosols comprise the submicron, <0.02 μ m (viruses, endotoxin, and mycotoxin), to multi-micron, 0.2–50 μ m (bacteria, fungi, parasites, and algae), biological particulates suspended in air, as live or dead intact microbes or their constituents/fragments, which may also include endotoxin, mycotoxins, insect parts, pollen, grain and microbial proteins (Cox and Wathes, 1995). Bioaerosols may be generated either as liquid droplets or as dry materials and transit in air either individually, as clusters, or on 'rafts' of organic matter. While bioaerosols are ubiquitous in ambient air and are carried short and long distances by small and large air currents (Brown

^{*} Tel.: +1 301 504 5631; fax: +1 301 504 8370. E-mail address: pat.millner@ars.usda.gov.

Mogens and Hovmøller, 2002; Jones and Harrison, 2004; Lighthart and Mohr, 1994), the types and amounts can vary considerably in space and time depending on several physico-chemical factors. In general, confined animal operations and the generation of bioaerosols, dust, and odors have accentuated the need for development of sustainable solutions to reduce adverse health impacts from these constituents on workers, animals, and nearby communities (Clark et al., 1983; Cole et al., 2000; Douwes et al., 2003; Nowak, 1998; Seedorf et al., 1998; Zejda et al., 1994).

In ambient indoor and outdoor air, except when a source is nearby, concentrations of potential public or veterinary pathogens generally are very low or absent. Several studies have been conducted to determine the extent to which bioaerosols generated at animal production facilities are significantly different from those in the general ambient air. Table 1 shows examples of the ambient airborne concentrations of cultured microorganisms reported from various animal confinement (indoor) situations in which reduction technologies have yet to be implemented.

2.1.1. Bioaerosol mitigation

More information is needed to understand and design air quality management technologies (indoor or exhaust systems) that are effective in reducing bioaerosols. Reports indicate that some systems function satisfactorily under standard operations for removal or control of contaminants like odor and particulates, but can fail to control emissions of certain pathogenic agents responsible for animal disease outbreaks. This has important implications for emergency preparedness, and animal disease epidemic management. For example, a dual bioscrubber-chemfilter system operating to reduce odor and ammonia emissions also significantly reduced emissions of airborne dust, endotoxins, and bacteria at a duck fattening facility (Zucker et al., 2005). However, Aarnink et al. (2005) reported that existing odor and NH₃ scrubbers, failed to prevent bioaerosol emissions during a swine fever outbreak in the Netherlands. They determined that supplementation of the sulfuric acid in existing scrubbers with peracetic acid in times of high risk of disease outbreak would be the most cost-effective way to prevent spread of the disease agent, because this more effective disinfectant would be impractical and cost prohibitive for continuous use.

2.2. Dust

Dust as fine particulates varies in shape, size, and composition, but can enter and deposit in the upper airways and deep lungs (AC-GIH, 2006). Deposition site depends on size and may range from inhalable ($\leq 100 \, \text{um}$ aerodynamic diameter (d_{ae})), thoracic (\leq 10 um d_{ae}) to respirable (\leq 5 um d_{ae}). Although fine particulates may be composed of inorganic or organic constituents, microorganisms (live or dead, whole or fragmented) may be enmeshed or attached to them. Such fine particulates may also have volatile compounds such as NH₃ or a range of organic compounds adsorbed to them (Bottcher et al., 2004; Schiffman et al., 2000; Schiffman and Williams, 2005). In people, long-term exposure to such dusts can cause respiratory damage, and in the short-term, effects can result from daily workshift exposure <1.0 mg/m⁻³ respirable dust. Symptoms in workers may include: bronchitis, hyperactive airways disease, atopic asthma, acute organic dust toxic syndrome, chronic organic dust toxic syndrome, mucous membrane irritation, increased susceptibility to other chest illnesses, chronic sinusitis, a byssinosis-like condition, nausea, diarrhea, rhinitis, fatigue, eye and throat irritation, headache, shortness of breath, wheezing, dizziness, and sleep disturbances. This extensive list of symptoms reflects the vast number of potential bioactive constituents in dusts.

From the perspective of occupational exposure in agricultural settings, endotoxin is probably the most relevant parameter so far identified with respirable particulates and associated with lung function impairment (Nowak, 1998; Spaan et al., 2006). Its microbial origin, ubiquity, persistence, and capacity to attach to substances and particulates makes it a challenging material to control in agricultural operations. As noted by Spaan et al. (2006), agricultural industries in general often exceed the Dutch proposed occupational exposure limit (50 endotoxin units-EU m⁻³) and temporary legal limit (200 EU m⁻³) for airborne endotoxin. The latter is a primary example of a material which is in part a bioaerosol (because it is microbially derived) and a dust (because

Table 1Bioaerosol concentrations indoors at various types of animal production facilities.

Microbes	Concentration (cfu/m³ air)	Sampler type	Reference
Total	1.5×10^{7}	Filtration	Larsson et al. (1988)
Swine Total Gram-negative	1.5×10^{5}	Impactor (An6 [*])	Cormier et al. (1990)
	80		
Fungi			
Total		Impactor (An6)	Heederik et al. (1991)
Gram-negative			
Total		Impinger, Filtration	Clark et al. (1983)
Gram-negative	8.8×10^4		
Fungi			
Total	7.7×10^{6}	Cyclone	Hinz and Linke (1998)
Gram-negative			
Total		Filtration, Impactor (An6) Filtration, Impactor (An6)	Predicala et al. (2002)
Respirable			
Total		Impactor (Slit)	Banhazi et al. (2005)
	2.17×10^{5}		
Total bacteria	1.3×10^{4}	Impactor (An1*)	Kim et al. (2008)
Total fungi	1.3×10^{3}		· ,
Total bacteria	$2.9-3.8 \times 10^4$	Impactor (SAS100**)	Pavicić et al. (2008)
Total coliform	$5.8-6.9 \times 10^2$		` ,
Hemolytic bacteria			
Fungi	$4.4-5.5 \times 10^2$		
	Total Total Gram-negative Fungi Total Gram-negative Total Gram-negative Fungi Total Gram-negative Fungi Total Gram-negative Total Respirable Total Total Total bacteria Total fungi Total bacteria Total coliform Hemolytic bacteria	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	$ \begin{array}{cccccccccccccccccccccccccccccccccccc$

^{*} An6, Andersen, 6-stage impactor; An1, Andersen, 1-stage impactor.

^{**} SAS100, the manufacturer and model of impactor.

of its size and nonviable nature). Seedorf et al. (1998) reported from a survey of cattle, pig, and poultry housing units in four European countries that cattle houses had the lowest endotoxin concentrations whereas poultry houses had the greatest with a mean of 692 and 49 ng/m³ for inhalable and respirable particulates, respectively.

Occupational standards specify threshold limit concentrations for airborne respirable dust and protection of workers (AGCIH, 2006) and standard methods for collection and measurement of respirable dust are available (NIOSH, 1994; Phillips et al., 1998). However, differences in collection efficiencies have been reported for several personal samplers in use, and work-specific conversion coefficients to normalize estimates of worker exposure to total dust from measurements using various dust sampling devices have been derived and suggested by Predicala and Maghirang (2003). In addition, O'Shaughnessy et al. (2007) provided a method for correcting a sampler-to-sampler ratio to deal with changes in size distribution of aerosols that can be used in most conditions, provided initial comparisons are normalized with the same dust.

2.2.1. Dust mitigation

In general, with adequate ventilation of animal housing, decreased stocking density, and structural and regular management of manure exposure to dust and endotoxin can be minimized (Banhazi et al., 2005; Duchaine et al., 2000). Canola oil spray reduced dust in swine barns and reduced acute health effects in naive healthy workers (Senthilselvan et al., 1997). Spraying rape seed oil daily was more effective at reducing aerial dust concentrations then adding 4% fat to the swine diet, resulting in 75% and 50% reductions in dust, respectively (Pedersen, 1998). Electrostatic space charge systems have shown promise in reducing dust in poultry houses (Ritz et al., 2006; Mitchell and Baumgartner, 2007). Similarly, electrostatic charge systems were also shown to reduce Salmonella spp. by 50% in chicks and in their ceca (Gast et al., 1999), while also reducing dust concentrations. As these and other mitigation technologies are developed and evaluated further, it will be important to determine their effects on the emissions of bioaerosols, pathogens and their viability/infectivity, endotoxin, and odorants.

2.3. Odorous volatile compounds

Odors are a perceived response (Schiffman and Williams, 2005) to certain volatile organic compounds (VOCs), but these odorants (the compounds) are not necessarily correlated with the presence or amounts of pathogens or fecal indicator microorganisms in the materials (Kim et al., 2005), particulate matter, or endotoxin. Human panels, olfactometry standards and practices, and GC–MS analysis (Schiffman et al., 2001; Zahn et al., 1997) coupled with sniffing ports have aided evaluation of the odor response in connection with animal facility odorant concentrations.

All animal operations generate odorous volatile compounds from microbial metabolism of the various organic materials present in the systems, including feed, bedding, and excreta. Many of these compounds are detectable at exceedingly low concentrations (Millner and McConnell, 2000; Ruth, 1986; Schiffman et al., 2000). Schiffman et al. (2001) reported 331 different VOCs and fixed gases from swine facilities. In confined interior spaces with intensive animal stocking densities, some odorous compound concentrations (NH3 and H2S), can accumulate rapidly and become a respiratory hazard for workers and animals. Beyond animal facility perimeters per se, however, odor complaints often lead to complaints about feeling ill (health symptoms), which in turn lead members of surrounding communities to express concerns about infection or toxicity from exposure to what may be transported in the air along with the odorants. Schiffman et al. (1995) have reported that these and other con-

cerns and responses to perceptions of odor are significantly associated with mental health consequences, such as increased tension, depression, fatigue, confusion, and mood changes. Such responses also match closely those reported for nuisance odors.

Schiffman and Williams (2005) described three mechanisms by which ambient odors might elicit health symptoms: (1) exposure to odorous compounds elicits a response in the trigeminal nerve system referred to as an irritation effect (irritation causes the health symptoms, whereas odor is the exposure marker); (2) at nonirritant concentrations exposure elicits an innate, learned aversion; (3) copollutants (such as endotoxin) elicit a health symptom response. They note that objective bio-markers of health symptoms are needed, to determine if and when health complaints constitute health effects. For these reasons, measurements of volatile compounds, dust (particulates which can absorb and re-volatilize compounds), odor, and irritation are needed along with measurements of bioaerosols and endotoxin to discern the contribution of each and the health effect. An additional factor that complicates the situation is that odorous compounds, even when individually present below their irritant threshold concentrations, can when present in mixtures collectively exceed an irritant threshold concentration and thus elicit a response in a sensitive receptor (Korpi et al., 1999).

2.3.1. Odor mitigation

As noted above, biofilters and chemical air scrubbers may be effective in reducing emissions of odorous compounds from animal production facilities. In general, with adequate ventilation and regular management of manure in animal housing, $\rm H_2S$ and $\rm NH_3$ may be maintained below 20 ppm and 10 ppm, respectively, and thereby avoid adverse respiratory responses to these hazards in workers and animals. However, effects of technologies like ventilation and practices on mitigating bioaerosols and endotoxin must be evaluated to determine the potential for health effects from chronic exposure to low concentrations.

Loughrin et al. (2006) reported substantial reductions (83–97%) in lagoon liquid odorants, *p*-cresol (83%), 4-ethylphenol (93%), and skatole (97%) resulting from a multi-stage swine manure treatment system. In studies of ambient air outside swine facilities, Lemay et al. (2007) found that NH₃ and H₂S are rapidly diluted well below toxic concentrations. Nonetheless, because adsorption of compounds to particulates that are highly respirable and exposure to other kinds of volatile organic compounds has been shown to influence immune response, Schiffman and Williams (2005) noted that more research is needed to evaluate these types of impacts on human health.

3. Bioaerosol collection and analyses in animal stables and farms

Microbial samplers available to collect bioaerosols include single and multi-stage impactors, impingers, filters, cyclones, vertical elutriators, and electrostatic precipitators. Details of each are described by Cox and Wathes (1995) and Henningson and Ahlberg (1994). Most have been used to assess exposure concentrations indoors in workplaces, homes, and schools and have been extended for use in outdoor air, although their efficiencies in outdoor settings, in which variable air speeds and directions occur, have not been well documented.

With specific regard to bioaerosols, there is a notable absence of standardized and validated methods for enumeration of various types of microorganisms in outdoor bioaerosols. Although various methods and devices have been used to detect and quantify outdoor bioaerosols, results show a wide range in values for prevalence and concentration across very diverse types of animal

operations and landscapes. Without doubt, standardization and validation of collection and sample analysis protocols are currently the most important technological factors that needs to be resolved with regard to outdoor bioaerosol investigations. Until a standardized, validated methods are established for each of the microbial types or groups (bacteria, fungi, viruses, protozoa, and endotoxin) in outdoor air, then studies will continue to be of value only on a comparative basis within the context of the factors tested at the same site. Cross-site comparisons and multi-institutional datasets which eventually may be needed for development of standards and practices or regulatory compliance will require standardized, validated methods, devices, and protocols. A major difficulty that needs to be confronted from the fact that no single sampling approach, either impaction, impingement, filtration, centrifugation, or passive, is suitable for all groups of microbes. Each microbial group, and even within groups, has inherent survival limits that need to be matched with the type of sampler and the stress that it imparts to the particles during the collection process. As molecular methods have advanced, there are increasingly greater opportunities and examples for the utilization of identification approaches in the presence, concentration, and survival of the microflora in air. Additional research and development is needed to advance beyond the traditional culture-based methods for detection and quantification.

Advanced devices that take advantage of culture-independent techniques are being developed to help overcome the time lag inherent in existing bioaerosol protocols (Agranovski et al., 2006). Bioaerosol concentration studies often show large variations in samples, even with indoor air in adjacent locations (Toivola et al., 2002). Consequently, bioaerosol study designs need to include quality assurance (duplicates and field blanks) for the specific microbial targets being measured. Methods for endotoxin determinations from air samples also have recently been reviewed and evaluated (Spaan et al., 2007).

Cultivation-independent surveys of small-subunit rRNA genes used to assess total bacterial and fungal bioaerosol constituents have revealed a greater species richness than traditional culturedependent methods, and in some cases show 100- to 1000-fold greater diversity (Brodie et al., 2007; Després et al., 2007; Nehme et al., 2008). Fluorochromes also offer an alternative sensitive tool for detection and culture-independent quantification that detects relationships to important atmospheric physical factors (temperature, rainfall, and UV light) to which culture-dependent factors are relatively insensitive (Chi and Li, 2007). Maron et al. (2005) using culture-independent analyses determined that the majority of the identified bacteria were soil or plant associated. Thus, they concluded that the bioaerosols originated from local sources. In contrast, results from molecular approaches to bioaerosol analyses led Fierer et al. (2008) to conclude that short-term temporal variability in bacterial diversity was associated with location in ambient outdoor air.

4. Dispersal and transport

4.1. Land application

Manure management and application on fields is accomplished in a wide variety of ways on different animal production facilities, including: lagoons, solids stacking, sprinkler irrigation of liquids with forced-projection or center-pivot equipment, injection of slurries, slurry irrigation from vehicle mounted spray nozzles, and land application of solids with box-beaters, side discharge units, or V-box spreaders (NRAES, 1994).

Land application of lagoon sludge (solids 6–13%) may only occur once a year or less, but is inherently an aerosol and odor gen-

erating process. The three main materials-handling approaches used for this liquid include: (1) pumping slurry through a large bore gun-sprinkler system on cropland followed by soil incorporation; (2) initial dewatering and subsequent manure spreading on fields; (3) dewatering and direct placement on cropland by spreaders equipped to handle slurries. Where injection can be used, aero-solization potential may be considerably reduced. In systems that separate solids from liquids for beneficial use (such as bedding materials, supplements to animal feed rations, composting, and soil amendments), separators can readily be enclosed to minimize aerosolization.

Boutin et al. (1988) conducted an extensive series of bioaerosol tests during land application of cattle and swine slurry with mobile and stationary farm equipment. They reported that total bacterial counts in air at the edge of the applied areas was >2000/m³, and no pathogenic bacteria were recovered. High pressure spray gun systems that discharge the liquids upwards results in substantial droplet size reduction and drift, whereas tank spreading reduced the spray area arc and drift, resulting in reduced airborne bacteria concentrations relative to the spray gun.

The physical characteristics of municipal wastewater treatment solids (biosolids) are similar to manures treated with flocculating agents to increase solids content. Results from biosolids land application studies suggest that microbes in such moist, sticky, viscous types of materials are attached closely to sludge particles, and that the density propels the particles to a rapid deposition on the land rather than keeping them suspended in long-distance off-site transport (Brooks et al., 2005).

For relatively dry materials, like poultry litter, where screens, separators, and box spreaders are used, bioaerosol generation will occur and may result in longer distance transport due to its lower density and propensity to finer particles. Depending on the material and its stability these operations may also be accompanied by the release of odorant compounds that are detectable in the parts-per-billion range. In these cases, odorants that reach neighboring communities will elicit concerns as described above for odors. Limited data are available by which to clearly assess the risk to all segments of the receptor communities. Recent information relative to asthmatic children (Hoopmann et al., 2006) and exposure to bioaerosols from livestock operations points to the need for further evaluation of this subgroup to bioaerosols, dust, and odorants. These same issues apply to land application of manures and other organic residuals (Gerba and Smith, 2005; Boutin, 1988).

4.2. Modeling

Major interest in airborne survival and transport of microorganisms arises in connection with public health pathogens and zoonotic diseases. The focus in such cases is on minimizing dissemination of infectious agents and avoiding disease in animals or humans (Donaldson et al., 2001; Gloster et al., 2003; Harvey et al., 2007). In an effort to enhance preparedness and epidemic management, atmospheric transport models have been developed and applied to specific microbial agents (Cannon and Garner, 1999; Mayer et al., 2008). Transport principles and factors applicable to fine particulates are equally useful with odorous volatiles and bioaerosols which are often released simultaneously from the same materials-handling operations. In the past, bioaerosol modeling has relied heavily on standard Gaussian dispersion models with modification to account for die-off of microbes during air transit as a result of exposure to UV radiation, temperature, and desiccation (Lighthart and Mohr, 1994). A die-off constant for E. coli was reported as $8.8 \times 10^{-3} \, \text{s}^{-1}$ for morning, and $6.6 \times 10^{-2} \, \text{s}^{-1}$ for afternoon (Teltsch et al., 1980). Clearly, using a resistant spore-former such as Clostridium spp., which would be expected to easily survive ambient atmospheric stressors, as a surrogate for off-site transport would provide a poor estimate of survival of non-spore formers, like *E. coli* and *Salmonella* spp., at distant downwind locations.

A variety of dispersion models, including Lagrangian, computational fluid dynamic, and modified Gaussian plume and puff models have been developed to deal with short and long range transport for epidemic management as well as odorous and pollutant compounds, and dust, from animal operations (Mayer et al., 2008; Minyoung et al., 2007; Nimmermark et al., 2005; Schiffman et al., 2003). For foot-and-mouth virus, local winds have a major influence on its dispersal and canalization of air flows in valleys accentuates the movement and concentrations (Mayer et al., 2008). With impinger collections from a center-pivot drop-nozzle irrigation system distributing swine lagoon liquid, Minyoung et al. (2007) reported increased total coliforms at 7.62 m downwind of the source, but at larger distances (70 m) concentrations were equivalent to those upwind. Wind direction variability during sampling periods coupled with the moving source presented unique sampling challenges. Similarly challenging is the situation with a tank truck moving while spraying liquid dairy slurry. In that case, impingers and high volume cyclone samplers were mounted on the front of an all-terrain vehicle which followed the tank truck at the downwind edge of the spray deposition. Total bacteria and coliform concentrations (2.5 \times 10³ and 3.5 \times 10²) were 100–1000 times more concentrated than upwind or the 50 and 200 m downwind samples (Millner, data unpublished), which is consistent with values reported by Boutin et al., 1988. In other studies, Köllner and Heller (2006) calculated using VDI guidelines (VDI, 2007) that total airborne bacterial concentrations downwind of animal farms in the Northern Rhine Westphalia region reached background density at a distance of 420 m from the farm. However, the greatest concentrations were found 50 and 75 m away from the facility.

An alternative approach to bioaerosol assessment as described by Paez-Rubio et al. (2006) relies on estimating aerosol source concentrations and emission rates by both reconstruction and correlations between bulk soil/solids mixture content and emission rates. They suggest that this approach would eliminate the need to directly measure pathogens or toxins in aerosols and thereby avoid difficulties and limitations associated with monitoring low aerosol concentrations of toxic compounds and pathogens. They applied this approach to estimating aerosol source concentrations and emissions rates with data on soils and biosolids and showed that disking presented the major biosolids-derived bioaerosol activity.

5. Summary

Quantifying the airborne constituents in the occupational environment by determining bioaerosols, dust, and odorants (volatile compounds) has important implications for evaluating the potential health risk to exposed people and animals and to animal productivity. Evaluating the exposure to known health hazards in the work environment allows identification of minimum achievable levels of exposure to the hazardous agent that are compatible with the suite of activities and management practices (new and old) in the production cycle. As new management practices are implemented, their effectiveness and impacts can be determined and possibly improved to meet target action levels. With land application of manure, clearly a liquid that has been treated to significantly reduce the number of fecal indicator microorganisms and/or pathogen concentration would be an example of a best available practice. Such treatment is unlikely to eliminate endotoxin. However, with unconstrained atmospheric dilution outdoors, maintaining elevated concentrations of temporary releases of endotoxin would be difficult. Odorants may remain a challenge for land application of unstabilized liquids in developed locations with residents nearby. Data on land spreading of poultry litter, which is very dry and prone to generation of particulates, and quantitative bioaerosol emission data could be used to determine if modifications in spreader equipment might reduce bioaerosols, endotoxin, and dust from this manure management operation.

Multi-analyte approaches to air emissions have emerged through the efforts of various research centers and collaborations (Hinz and Linke, 1998; Phillips et al., 1998; Seedorf et al., 1998; Vanotti et al., 2007). Continued work within a framework that targets analyses of all these major airborne constituents that contribute to the air quality of workers, animals, and the community, is needed to support the evaluation, design and improvement of modern animal production systems that are increasingly situated near communities.

6. Conclusions

Future bioaerosol studies of animal operations need to emphasize:

- Standardization/validation of collection methods, appropriate for microorganisms of concern, including indoor and outdoor settings.
- (2) Standardization/validation of analytical techniques, using molecular and advanced rapid technologies appropriate to target microorganisms.
- (3) Evaluation of effects of new mitigation technologies during and after their development on bioaerosols, dust, endotoxin, and VOC concentrations, using standardized/validated collection/analytical techniques, along with odor assessments, and emphasis on health impacts on animals and their growth and on susceptible individuals such as asthmatic children, elderly persons, and those with respiratory and immune deficiencies in communities near animal operations.

References

- Aarnink, A.J.A., Landman, W.J.M., Melse, R.W., Huynh, T.T.T., 2005. Systems for eliminating pathogens from exhaust air of animal house. Livestock environment VII. In: Proc. Seventh Int. Sympos., pp. 239–244.
- ACGIH, 2006. Threshold limit values and biological exposure indices. In: American Conference of Governmental Industrial Hygienists ACGIH, Cincinnati, Ohio.
- Agranovski, I.E., Safatov, A.S., Sergeev, A.A., Pyankov, O.V., Petrishchenko, V.A., Mikheev, M.V., Sergeev, A.N., 2006. Rapid detection of airborne viruses by personal bioaerosol sampler combined with the PCR device. Atmos. Environ. 40, 3924–3929.
- Banhazi, T.M., Seedorf, J., Rutley, D.L., Pitchford, W.S., 2005. Statistical modeling of airborne bacteria and endotoxins concentrations in australian piggery buildings. Livestock environment VII. In: Proc. Seventh Int. Sympos., pp. 72–78.
- Bottcher, R.W., Keener, K.M., Munilla, R.D., Williams, C.M., Schiffman, S.S., 2004. Dust and odor emissions from tunnel ventilated swine buildings in North Carolina and comparison of different odor evaluation methods. Appl. Eng. Agric. 20, 343–347.
- Boutin, P., Torre, M., Serceau, R., Rideau, P.-J., 1988. Atmospheric bacterial contamination from landspreading of animal wastes: evaluation of the respiratory risk for people nearby. J. Agric. Eng. Res. 39, 149–160.
- Brodie, E.L., DeSantis, T.Z., Parker, J.P.M., Zubietta, I.X., Piceno, Y.M., Andersen, G.L., 2007. Urban aerosols harbor diverse and dynamic bacterial populations. Proc. Nat. Acad. Sci. 104, 299–304.
- Brooks, J.P., Tanner, B.D., Gerba, C.P., Haas, C.N., Pepper, I.L., 2005. Estimation of bioaerosol risk of infection to residents adjacent to a land applied biosolids site using an empirically derived transport model. J. Appl. Microbiol. 98, 397–405.
- Brown Mogens, J.K.M., Hovmøller, S., 2002. Aerial dispersal of pathogens on the global and continental scales and its impact on plant disease. Science 297, 537–541.
- Cannon, R.M., Garner, M.G., 1999. Assessing the risk of wind-borne spread of footand-mouth disease in Australia. Environ. Int. 25, 713–723.
- Chi, M.-C., Li, C.-S., 2007. Fluorochrome in monitoring atmospheric bioaerosols and correlations with meteorological factors and air pollutants. Aerosol Sci. Technol. 41, 672–678.
- Clark, S., Rylander, R., Larsson, L., 1983. Airborne bacteria, endotoxin and fungi in dust in poultry and swine confinement buildings. Am. Ind. Hyg. Assoc. J. 44, 537–541.

- Cole, D., Todd, L., Wing, S., 2000. Concentrated swine feeding operations and public health: a review of occupational and community health effects. Environ. Health Perspec. 108, 685–699.
- Cormier, Y., Tremblay, G., Meriaux, A., Brochu, G., Lavoie, J., 1990. Airborne microbial contents in two types of swine confinement buildings in Quebec. Am. Ind. Hyg. Assoc. J. 51, 304–309.
- Cox, C.S., Wathes, C.M., 1995. Bioaerosols Handbook. CRC Press, Boca Raton, Florida. Després, V.R., Nowoisky, J.F., Klose, M., Conrad, R., Andreae, M.O., 2007. Characterization of primary biogenic aerosol particles in urban, rural, and high-alpine air by DNA sequence and restriction fragment analysis of ribosomal RNA genes. Biogeosciences 4, 1127–1141.
- Donaldson, A.I., Alexandersen, S., Sørensen, J.H., Mikkelsen, T., 2001. Relative risks of the uncontrollable (airborne) spread of FMD by different species. Vet. Rec. 148, 602–604.
- Douwes, J., Thorne, P., Pearce, N., Heederik, D., 2003. Bioaerosol health effects and exposure assessment: progress and prospects. Ann. Occup. Hyg. 47, 187–200.
- Duchaine, C., Grimard, Y., Cormier, Y., 2000. Influence of building maintenance, environmental factors, and seasons on airborne contaminants of swine confinement buildings. Am. Ind. Hyg. Assoc. J. 61, 56–63.
- Fierer, N., Liu, Z., Rodríguez-Hernández, M., Knight, R., Henn, M., Hernandez, M.T., 2008. Short-term temporal variability in airborne bacterial and fungal populations. Appl. Environ. Microbiol. 74, 200–207.
- Gast, R.K., Mitchell, B.W., Holt, P.S., 1999. Application of negative air ionization for reducing experimental airborne transmission of *Salmonella enteritidis* to chicks. Poultry Sci. 78, 57–61.
- Gerba, C.P., Smith Jr., J.E., 2005. Sources of pathogenic microorganisms and their fate during land application of wastes. J. Environ. Qual. 34, 42–48.
- Gloster, J., Champion, H.J., Sørensen, J.H., Mikkelsen, T., Ryall, D.B., Astrup, P., Alexandersen, S., Donaldson, A.I., 2003. Airborne transmission of foot-and-mouth disease virus from Burnside Farm, Heddon-on-the-Wall, Northumberland, during the 2001 epidemic in the United Kingdom. Vet. Rec. 152, 525-533.
- Harvey, N., Reeves, A., Schoenbaum, M.A., Zagmutt-Vergara, F.J., Dubé, C., Hill, A.E., Corso, B.A., Bruce McNab, W., Cartwright, C.I., Salman, M.D., 2007. The North American animal disease spread model: a simulation model to assist decision making in evaluating animal disease incursions. Prev. Vet. Med. 82, 176–197.
- Heederik, D., Brouwer, R., Biersteker, K., Boleij, J.S.M., 1991. Relationship of airborne endotoxins and bacteria levels in pig farms with the lung function and respiratory symptoms of farmers. Int. Arch. Occup. Environ. Health 62, 595–601.
- Henningson, E.W., Ahlberg, M.S., 1994. Evaluation of microbiological air samplers: a review. J. Aerosol Sci. 25, 1459–1492.
- Hinz, T., Linke, S., 1998. A comprehensive experimental study of aerial pollutants in and emissions from livestock buildings. Part 2: results. J. Agric. Eng. Res. 70, 119–129.
- Hoopmann, M., Hehl, O., Neisel, F., Werfel, T., 2006. Associations between bioaerosols coming from livestock facilities and asthmatic symptoms in children [Zusammenhang zwischen bioaerosolen aus tierhaltungsanlagen und asthmatischen symptomen bei kindern]. Gesundheitswesen 68, 575–584.
- Jones, A.M., Harrison, R.M., 2004. The effects of meteorological factors on atmospheric bioaerosol concentrations a review. Sci. Total Environ. 326, 151–180.
- Kim, H., McConnell, L.L., Millner, P., 2005. Characterization of odorants from products of 14 different commercial composts using solid phase microextraction. Trans. ASAE 48. 315–320.
- Kim, K.Y., Ko, H.J., Kim, H.T., Kim, C.N., Kim, Y.S., 2008. Assessment of airborne bacteria and fungi in pig buildings in Korea. Biosys. Eng. 99, 565–572.
- Köllner, B., Heller, D., 2006. Ambient air concentrations of bioaerosols in the vicinity of a pigpen results of the project "health-related effects of bioaerosols emitted by livestock husbandries" | [Bioaerosolimmissionen im umfeld eines schweinemastbetriebes Ergebnisse aus dem projekt "gesundheitliche wirkungen von stall-luft-komponenten aus tierhaltungsbetrieben"]. Gefahrst. Reinhalt. Luft 66, 349–354.
- Korpi, A., Kasanen, J.P., Alarie, Y., Kosma, V.M., Pasanen, A.L., 1999. Sensory irritating potency of some microbial volatile organic compounds (MVOCs) and a mixture of five MVOCs. Arch. Environ. Health 54, 347–352.
- Larsson, K., Malmberg, P., Eklund, A., Belin, L., Blaschke, E., 1988. Exposure to microorganisms, airway inflammatory changes and immune reactions in asymptomatic dairy farmers. Int. Arch. Allerg. Appl. Immunol. 87, 127–133.
- Lemay, S.P., Godbout, S., Pelletier, F., Larouche, J.P., Belzile, M., 2007. Swine production impact on ambient gas and odor concentrations in agricultural areas. In: 2007 ASABE Ann. Int. Meeting, Technical Paper No. 074093. vol. 9, pp. 1–21.
- Lighthart, B., Mohr, A.J. (Eds.), 1994. Atmospheric Microbial Aerosols, Theory and Applications. Chapman and Hall, New York.
- Loughrin, J.H., Szogi, A.A., Vanotti, M.B., 2006. Reduction of malodorous compounds from a treated swine anaerobic lagoon. J. Environ. Qual. 34, 194–199.
- Maron, P.-A., Lejon, D.P.H., Carvalho, E., Bizet, K., Lemanceau, P., Ranjard, L., Mougel, C., 2005. Assessing genetic structure and diversity of airborne bacterial communities by DNA fingerprinting and 16S rDNA clone library. Atmos. Environ. 39, 3687–3695.
- Mayer, D., Reiczigel, J., Rubel, F., 2008. A Lagrangian particle model to predict the airborne spread of foot-and-mouth disease virus. Atmos. Environ. 42, 466–479.
- Millner, P., McConnell, L., 2000. Odor and other air quality issues associated with organic and inorganic by-products. In: Dick, W.A. (Ed.), Land Application of Agricultural, Industrial, and Municipal By-Products-SSSA Book Series No. 6, Madison, WI, pp. 289–314.

- Minyoung, K., Thurston, J.A., Hagen, L.J., 2007. Computational Fluid Dynamics (CFD) modeling to predict bioaerosol transport behavior during center pivot wastewater irrigation. In: Am. Soc. Agric. Biolog. Eng. Ann. Int. Meeting, Technical Paper No. 074063, vol. 8, pp. 1–8.
- Mitchell, B.W., Baumgartner, J.W., 2007. Electrostatic space charge systems for dust reduction in animal housing. In: 2007 ASABE Ann. Int. Meeting, Technical Paper number 074176, vol. 10, p. 13.
- Nehme, B., Létourneau, V., Forster, R.J., Veillette, M., Duchaine, C., 2008. Culture-independent approach of the bacterial bioaerosol diversity in the standard swine confinement buildings, and assessment of the seasonal effect. Environ. Microbiol. 10, 665–675.
- Nimmermark, S.A., Jacobson, L.D., Schmidt, D.R., Gay, S.W., 2005. Predictions by the odor from feedlots, setback estimation tool (OFFSET) compared with observations by neighborhood monitors. J. Air Waste Manage. Assoc. 55, 1306–1314.
- NIOSH, 1994. NIOSH Manual of Analytical Methods, fourth ed. National Institute for Occupational Safety and Health, NIOSH, Cincinnati, Ohio.
- Nowak, D., 1998. Health effects of airborne pollutants particularly in swine confinement houses a review focusing on aspects of occupational medicine [Die wirkung von stalluftbestandteilen, insbesondere in schweinestallen, aus arbeitsmedizinischer sicht]Dtsch. Tierarztl. Wochenschr. 105, 225–234.
- NRAES (Northeast Regional Agricultural Engineering Service), 1994. Liquid Manure Application Systems. NRAES-79. Northeast Regional Agricultural Engineering Service Coop. Exten. 152 Riley-Robb Hall, Cornell University, Ithaca, NY, USA, 215p.
- O'Shaughnessy, P.T., Lo, J., Golla, V., Nakatsu, J., Tillery, M.I., Reynolds, S., 2007. Correction of sampler-to-sampler comparisons based on aerosol size distribution. J. Occup. Environ. Hyg. 4, 237–245.
- Paez-Rubio, T., Hua, X., Anderson, J., Peccia, J., 2006. Particulate matter composition and emission rates from the disk incorporation of class B biosolids into soil. Atmos. Environ. 40, 7034–7045.
- Pavicić, Z., Balenović, T., Kabalin, A.E., Matković, K., Popović, M., Biuk-Rudan, N., Potocnjak, D., Gracner, G.G., 2008. Influence of number of piglets per unit of space on microbiological quality of air in rearing unit. Tierarztl. Umschau 63, 30–35
- Pedersen, S., 1998. Dust reduction in pig houses. Dtsch. Tierarztl. Wochenschr. 105, 247–250
- Phillips, V.R., Holden, M.R., Sneath, R.W., Short, J.L., White, R.P., Hartung, J., Seedorf, J., Schroder, M., Linkert, K.H., Pedersen, S., Takai, H., Johnsen, J.O., Groot Koerkamp, P.W.G., Uenk, G.H., Scholtens, R., Metz, J.H.M., Wathes, C.M., 1998. The development of robust methods for measuring concentrations and emission rates of gaseous and particulate air pollutants in livestock buildings. J. Agric. Eng. Res. 70, 11–24.
- Predicala, B.Z., Maghirang, R.G., 2003. Field comparison of inhalable and total dust samplers for assessing airborne dust in swine confinement barns. Appl. Occup. Environ. Hyg. 18, 694–701.
- Predicala, B.Z., Urban, J.E., Maghirang, R.G., Jerez, S.B., Goodband, R.D., 2002. Assessment of bioaerosols in swine barns by filtration and impaction. Curr. Microbiol. 44, 136–140.
- Ritz, C.W., Mitchell, B.W., Fairchild, B.D., Czarick III, M., Worley, J.W., 2006. Improving in-house air quality in broiler production facilities using an electrostatic space charge system. J. Appl. Poultry Res. 15, 333–340.
- Ruth, J.H., 1986. Odor thresholds and irritation levels of several chemical substances: a review. Am. Ind. Hyg. Assoc. J. 47, A-142–A-151.
- Schiffman, S., Williams, M., 2005. Science of odor as a potential health issue. J. Environ. Qual. 34, 129–138.
- Schiffman, S.S., Miller, E.A.S., Suggs, M.S., Graham, B.G., 1995. The effect of environmental odors emanating from commercial swine operations on the mood of nearby residents. Brain Res. Bull. 37, 369–375.
- Schiffman, S.S., Walker, J.M., Dalton, P., Lorig, T.S., Raymer, J.H., Shusterman, D., Williams, C.M., 2000. Potential health effects odor from animal operations, wastewater treatment, and recycling of byproducts. J. Agromed. 7, 7–81.
- Schiffman, S.S., Bennett, J.L., Raymer, J.H., 2001. Quantification of odors and odorants from swine operations in North Carolina. Agric. Forest Meteorol. 108, 213–240.
- Schiffman, S.S., Graham, B., McLaughlin, B., Fitzpatrick, D., Katul, G., Nagle, T., Williams, C.M., 2003. Predicting odor dispersion at five swine facilities using a Eulerian–Lagrangian model. In: Havenstein, G.B. (Ed.), Proc. North Carolina Animal Waste Manage. Workshop, Research Triangle Park, NC. 16–17 October 2003. North Carolina State Univ. College of Agric. and Life Sci., Raleigh, pp. 102–111.
- Seedorf, J., Hartung, J., Schroder, M., Linkert, K.H., Phillips, V.R., Holden, M.R., Sneath, R.W., Short, J.L., White, R.P., Pedersen, S., Takai, H., Johnsen, J.O., Metz, J.H.M., Groot Koerkamp, P.W.G., Uenk, G.H., Wathes, C.M., 1998. Concentrations and emissions of airborne endotoxins and microorganisms in livestock buildings in Northern Europe. J. Agric. Eng. Res. 70, 97–109.
- Senthilselvan, A., Zhang, Y., Dosman, J.A., Barber, E.M., Holfeld, L.E., Kirychuk, S.P., Cormier, Y., Hurst, T.S., Rhodes, C.S., 1997. Positive human health effects of dust suppression with canola oil in swine barns. Am. J. Respir. Crit. Care Med. 156, 410–417.
- Smith, J.E., Jr., Millner, P.D., Jakubowski, W., Goldstein, N., Rynk, R. (Eds.), 2005. Contemporary Perspectives on Infectious Disease Agents in Sewage Sludge and Manure. JG Press Inc., Emmaus, PA, p. 249pp.
- Spaan, S., Wouters, I.M., Oosting, I., Doekes, G., Heederik, D., 2006. Exposure to inhalable dust and endotoxins in agricultural industries. J. Environ. Monit. 8, 63–72.

- Spaan, S., Heederik, D.J.J., Thorne, P.S., Wouters, I.M., 2007. Optimization of airborne endotoxin exposure assessment: effects of filter type, transport conditions, extraction solutions, and storage of samples and extracts. Appl. Environ. Microbiol. 73, 6134–6143.
- Strauch, D., Ballarini, G., 1994. Hygienic aspects of the production and agricultural use of animal wastes. J. Vet. Med. B 41, 176–228.
- Teltsch, B., Shuval, H.I., Tadmor, J., 1980. Die-away kinetics of aerosolized bacteria from sprinkler application of wastewater. Appl. Environ. Microbiol. 39, 1191–1197.
- Toivola, M., Alm, S., Reponen, T., Kolari, S., Nevalainen, A., 2002. Personal exposures and microenvironmental concentrations of particles and bioaerosols. J. Environ. Monit. 4, 166–174.
- Vanotti, M.B., Szogi, A.A., Hunt, P.G., Millner, P.D., Humenik, F.J., 2007. Development of environmentally superior treatment system to replace anaerobic swine lagoons in the USA. Bioresour. Technol. 98, 3184–3194.
- VDI (Verein Deutscher Ingenieure, The Association of Engineers), 2007. Technical Rule, 2007-02, 4251 Blatt 1. Measurement of Airborne Microorganisms and Viruses in Ambient Air – Planning of Plant-Related Ambient Air Measurements
 Plume measurement. Kommission Reinhaltung der Luft (KRdL) im VDI und DIN – Normenausschuss (Commission on Air Pollution Prevention of VDI and DIN – Standards Committee). Beuth Verlag GmbH, D-10772 Berlin, Germany.
- Zahn, J.A., Hatfield, J.L., Do, Y.S., DiSpirito, A.A., Laird, D.A., Pfeiffer, R.L., 1997. Characterization of volatile organic emissions and wastes from a swine production facility. J. Environ. Qual. 26, 1687–1696.
- Zejda, J.E., Barber, E.M., Dosman, J.A., Olenchock, S.A., McDuffie, H.H., Rhodes, C.S., Hurst, T.S., 1994. Respiratory health status in swine producers relates to endotoxin exposure in the presence of low dust levels. J. Occup. Med. 36, 49–56.
- Zucker, B.-A., Scharf, P., Kersten, C., Müller, W., 2005. Influence of an exhaust air washer on the emission of bioaerosols from a duck fattening unit. Gefahrst. Reinhalt. Luft. 65, 370–373.